

STAT-6 In Hodgkin Lymphoma Pathobiology and Treatment-Review of The Literature

M Ioannou*, K Baxevanidou, GK Koukoulis

Department of Pathology, University of Thessaly, Greece

Volume 1 Issue 5- 2018

Received Date: 01 Aug 2018

Accepted Date: 20 Aug 2018

Published Date: 27 Aug 2018

2. Keywords

Hodgkin lymphoma; STAT6; Pathobiology; Therapy; Review

1. Abstract

Classical Hodgkin Lymphoma (cHL), consists of rare neoplastic Hodgkin and Reed-Sternberg cells (HRS) residing in a prominent inflammatory background. HRS show deregulated activation of multiple signaling pathways and transcription factors. The activation of these pathways and factors is partly mediated through interactions of HRS with various other types of cells in the microenvironment, but also through genetic lesions. Signal transducers and activators of transcription (STAT) are a family of transcription factors that regulate a broad range of cellular processes, such as proliferation, differentiation, and survival, in a large variety of cell types. STAT6 pathway is activated as a response to the binding of cytokines IL-4 and IL-13 to their receptors on the cell membrane. The ability of activated STAT6 to promote lymphoproliferation and the requirement for STAT6 in normal cytokine-induced cell proliferation provides a strong rationale for further study of STAT6 in cHL.

This review outlines the current evidence on the role of STAT6 in cHL. We report on the findings concerning the involvement of STAT6 in the pathogenesis, as well as in the cross-talk between tumor cells and their microenvironment. The dependency of HRS on micro environmental interactions and on deregulated STAT6 signaling pathway may offer novel strategies for targeted therapies.

3. Introduction

Hodgkin lymphoma (HL) was recognized in the first half of the 19th century by Thomas Hodgkin and Samuel Wilks [1,2]. It is one of the most common lymphomas in Western World. Its annual incidence is 3 cases per 100.000 persons. Neoplastic tissues usually contain a small number of scattered large mononucleated and multinucleated tumor cells (designated Hodgkin and Reed-Sternberg cells or HRS cells) residing in an abundant heterogeneous admixture of non-neoplastic inflammatory and accessory cells. The latter includes lymphocytes, especially Th2 cells, monocytes, granulocytes, eosinophils, mast cells and histiocytes [1,3,4]. Biological and clinical studies in the last decades have shown that Hodgkin lymphomas are comprised of two disease entities: nodular lymphocyte predominant Hodgkin lymphoma (NLPHL) and classical Hodgkin lymphoma (cHL). Based on the consistence of the microenvironment, the latter one is divided into four subtypes: nodular sclerosis (80%), mixed cellularity (15%), lymphocyte rich (5%) and lymphocyte depleted (<1%) (1, 5, 6). The immunophenotypic and genetic

features of the mononuclear and multinucleated cells are identical in these histological subtypes, whereas their clinical features and association with EBV show differences.

In Epstein-Barr virus (EBV) positive cases evidence suggest its role in the pathogenesis of HRS cells [5,7-11]. The prevalence of EBV in HRS cells varies according to the histological subtype and epidemiologic factors. The highest frequency is found in mixed cellularity classical HL and the lower incidence in nodular sclerosis classical HL (WHO 2008). EBV is found in 40% of cases in Western world, however may be seen in up to 90% of cases in Central and South Africa [5].

Classical HL is a monoclonal lymphoid neoplasm derived (in most instances from B cells). Despite their derivation from germinal center B cells, HRS have lost much of the B-cell specific expression program and have acquired B-cell inappropriate gene products. In addition, deregulated transcription factors in classical HL promote proliferation and abrogate apoptosis in the neoplastic cells. Multiple signaling pathways, mainly including nu-

*Corresponding Author (s): M Ioannou, University of Thessaly, School of Health Sciences, Department of Pathology, Panepistimion 3st Biopolis, 41110, Larissa, Greece, E-mail: mioan@uth.gr

clear factor kappa-B (NF- κ B), Janus kinase-signal transducer and activator of transcription signaling (JAK/STAT), PI3K-Akt and ERK, have deregulated activity in HRS cells. Coherently, recurrent genetic alterations detected in HRS cells of cHL frequently affect members of the NF- κ B or JAK/STAT signaling pathways [5,8,10,12]. CHL is associated with over expression and an abnormal pattern of cytokines and chemokines including IL-5, IL-6, IL-7, IL-9, IL-10, IL-13 and granulocyte-macrophage colony-stimulating factor and/or their receptors in HRS cells [13-17] which likely explain the abundant admixture of inflammatory cells, fibrosis and the predominance of Th2 cells in the infiltrating T-cell population [18].

STAT-6 is a protein that belongs to a family of transcription factors known as STATs. STAT6 is one of the seven members of STAT protein family (STAT1, STAT2, STAT3, STAT4, STAT5a, STAT5b, STAT6), which was identified and cloned for the first time by two independent teams [19]. The signaling cascade of the JAK-STAT pathway is triggered by the engagement of cytokine receptors. This leads to the activation of docking domains for STAT monomers. After having bounded to the phosphotyrosines of their receptors the STATs themselves are being phosphorylated on tyrosine residues, which enables them to form dimers. The active STAT dimers translocate to the nucleus, where they control the expression of target genes [20].

There are only few publications regarding STAT6 in HL. STAT6 activation in cHL is mediated by IL-13, and this has been proved by immunohistochemistry, immunoblotting, Western blotting and ELISA. Notably, co-expression of IL-13 and its receptor IL-13R α 1 is characteristic for HRS cells [21-24].

The current treatment of HL is based on multi-drug chemotherapy, radiation therapy (25, 26) and autologous or allogenic stem cell transplantation in case of recurrence [26]. Recent studies provided insights into deregulation of key nodal signaling pathways, including the PI3K [27], NF- κ B [28-30] and JAK/STAT [31] pathways, which are amenable to small-molecule targeting. Understanding how neoplastic cells interact and depend on their microenvironment has led to a remarkable new step in developing new treatment strategies targeting not only the malignant tumor cells but also the tumor microenvironment [25,26].

In this context, the present review outlines the current evidence on the role of STAT6 in cHL. Molecular, and histopathological data regarding STAT6 expression in neoplastic cells are presented along with the possible involvement in pathogenesis, and pathobiology of cHL. In addition, the current evidence for its potential use as a therapeutic target is discussed.

4. STAT-6

The signal transducers and activators of transcription (STATs) including STAT6 are latent cytoplasmic proteins that undergo tyrosine phosphorylation by Janus kinases (JAKs) in response to cytokine exposure in the extracellular milieu. Ligation of cytokines interleukin-4 (IL-4) and interleukin-13 (IL-13) with their receptors result in a common STAT6-mediated signaling pathway critical for a number of responses in T cells, including the development of T helper type 2 (Th2) cells and IL-4-stimulated proliferative responses, functions that were demonstrated through the analysis of mice with disrupted Stat6 alleles [32-34]. Once phosphorylated, STAT6 is transported to the nucleus where it regulates gene expression in various cell types critical to the balance between host immune defense and inflammatory responses [35,36]. IL-4, IL-13, and STAT6 promote humoral immunity, clearance of helminthic parasites as well as the pathogenesis of allergic disorders like asthma, food allergies, and atopic dermatitis [37-39].

While STAT6 is required for normal immune function, it has been also implicated in numerous malignancies including prostate and colon cancer [40-42], glioblastoma [43], lymphoma [21,44,45], and leukemia [46,47]. A recurrent intra-chromosomal rearrangement on chromosome 12q leading to the formation of a NAB2-STAT6 fusion oncogene has been recently identified in solitary fibrous tumor [48,49]. As a result, nuclear expression of the cytoplasmic transcription factor STAT6 is found in solitary fibrous tumor and serves as a useful diagnostic marker [50, 51]. Using Fluorescence In Situ Hybridization (FISH) analysis, Doyle et al detected STAT6 amplification in a subset of dedifferentiated liposarcoma with nuclear immunorexpression of STAT6 protein [52]. STAT6 amplification has also been demonstrated in Hodgkin lymphoma cell lines [53]. Currently, experimental therapeutics that target the IL-4/IL-13/STAT6 pathway are being tested in clinical trials [54, 55]. The involvement of STAT6 in human oncogenesis is opening up new possibilities regarding the study and identification of new molecular targets for the development of future cancer therapy.

5. Stat6 and Hodgkin Lymphoma

5.1. Stat6 In Pathogenesis and Tumor Growth

The malignant cells in cHL (HRS cells) arise from the germinal center cells or the immediate post germinal center cells. Although originating from B-lymphoid cells, HRS cells have lost their B cell-phenotype and show co-expression of markers characteristic for other hematopoietic lineages [5,56]. HRS cells are characterized by the constitutive activation of nuclear factor kappa B (NF- κ B), the Activator Protein-1 (AP-1), the

deregulation of lineage-specific transcription factors such as E2A [5], and the Interferon regulating factor (IRF)5 that, together with NF- κ B activation, determine the inflammatory phenotype of HRS cells [57]. HRS cells express CD30 and CD40, two members of the tumor necrosis factor (TNF)/nerve growth factor (NGF) receptor family, and in the majority of cases CD15 (75–85%) and IRF4 [5].

The JAK/STAT signaling pathway represents another key pathway in pathogenesis of HL. STAT3, STAT5 and STAT6 are activated and expressed at high level in HL [21, 58]. Given that the activation of docking domains for STAT monomers is due the activation of JAK/STAT pathway, the expression of STATs and especially of STAT6 in HRS cells might represent a possible biomarker of JAK/STAT activation in tissue specimens. Clinicopathological studies correlating clinical data and molecular results with immunohistochemical expression of STAT6 protein, could further investigate this possibility.

Epstein–Barr virus (EBV) is causally associated with approximately one third of HL cases in socioeconomically developed countries, while in pediatric HL in Central and South America, the association can be up to 90% [59]. In patients with AIDS, EBV-infected HRS cells are present in nearly all cases [60]. Different studies have shown that Epstein-BarrVirus contributes to the transformation of its precursors, as well as the survival and proliferation of the malignant HRS cells [4,5,7]. The EBV+ HRS cells typically show an EBV latency II gene expression profile, meaning expression of the viral proteins EBV nuclear antigen 1 (EBNA1) and latent membrane proteins 1 and 2a (LMP1 and LMP2a) [61]. The EBV-encoded LMP-1 is a viral mimic of the CD40 receptor, and by constitutive signaling it activates potently the nuclear factor κ B, c-Jun N-terminal kinase, and phosphatidylinositol 3-kinase pathways. LMP-1 has been reported as a viral oncoprotein promoting tumor growth but also apoptotic resistance and immune modulation [9,62,63]. Recently, demonstrated that the induction of LMP-1 by IL-4 and IL-13 is mediated by STAT6 and a newly defined high-affinity STAT6-binding site in the LMP-1 promoter in HL-derived, EBV-converted KMH2-EBV cell lines [10]. This evidence strongly supports the role of STAT6 in the pathogenesis of EBV-positive cHL. Furthermore, it indicates that inhibition of the interactions between the cytokines and its specific receptor or inhibition of the STAT6 signaling pathway might have beneficial effects in the EBV-positive cases by down-regulating the expression of LMP-1.

In respect to tumor growth, it has been proved that survival and proliferation of HRS cells is dependent on STAT3 and STAT6 activation, since rescission of their activation by neutralizing antibodies, JAK/STAT blockers or siRNAs against STAT3 and STAT6 reduced proliferation and induced cell death in vitro [14, 25, 64]. In addition, pointed out the association between antibody-mediated

neutralization of IL-13, reduced STAT6 phosphorylation and decreased HL cell proliferation [19].

Interestingly, Baus et al. [55] demonstrated that knockdown of STAT6, by using constantly expressed shRNAs against STAT6 by lentiviral transduction, induced apoptotic cell death in the cHL cell lines L428 and L1236cHL. In the latter study, the values of the G1 and G2 cell populations were not affected, suggesting that STAT6 has a strong effect on cell survival and does not provoke cell-cycle arrest.

The above data suggest that STAT6 promotes the neoplastic proliferation and consequently it might represent a possible therapeutic target.

5.2 Stat6 and HL Microenvironment

Unlike any other neoplasm, the tumor in cHL is made predominantly of the non-neoplastic HRS cells rather than the neoplastic HRS cells, which often constitute no more than 1-3% of the entire mass [12]. CD4+ T cell lymphocytes are the most abundant cell type in cHL, clustering around the RS cells [48], and the overproduction of helper T cell type 2 (Th2) cytokines and chemokines such as interleukin (IL)-13, IL5 and eotaxin [14] is reported in most cases. The great number of cytokines produced in cHL by HRS cells promote neoplastic cell growth and survival. At the same time, the secreted molecules are implicated in the reactions between the cells of microenvironment and trigger an abnormal immune response to the HRS cells while support them to overcome the antitumor activity of cytotoxic T and NK cells [8]. This is highlighted by expression or secretion of PD-1 ligand, galectin-1 and IL-13 which directly interfere with the functional activity of T cells, primarily polarize specific T cell subsets towards a regulatory phenotype, or prevent an effective Th1-response [65-68].

Early studies reported that IL-13 and IL-13R (alpha)1, the IL-13-specific receptor chain, are frequently expressed by HL-derived cell lines as well as by HRS cells from biopsy material of tissues involved by HL. Furthermore, antibody-mediated neutralization of IL-13 in cultures of HL-derived cell lines resulted in a dose-dependent inhibition of proliferation, and it was associated with increased apoptosis and with significant decreases in both cellular proliferation and levels of phosphorylated STAT6 of HL cell lines [13, 14, 69].

These data support the hypothesis that STAT6 is involved in cellular interactions that modify the tumor microenvironment which, on the other hand, regulate the immune response against tumor cells. Since HRS survival seems to be mostly due the activated proliferating and proinflammatory cytokine secreting cells [8, 70], the IL-13/STAT6 signaling, involved in microenvironment

tal interactions, may be an additional target for new therapeutic approaches.

5.3. Stat6 and Therapy of HL

The majority of patients with HL are treated with a combination of multi-drug chemotherapy and radiotherapy. Despite relative success of therapy, approximately 20% of patients will not be cured with the current available therapy and the disease will relapse [26]. Moreover, 30-35% of patients with high-risk prognostic features will not be treated [71]. Additionally, patients recurring after autologous and/or allogenic stem cell transplantation are regarded incurable and are considered to have a median survival <3 years [72]. Hence, the development of novel therapeutic agents are needed for patients with refractory or relapsed disease.

Novel therapeutic strategies focus on the special and unique pathology and microenvironment, the deregulated signaling pathways, as well as the induction of anti-HRS cell immunity by modulating the microenvironment. Among the latter, the immune checkpoint inhibitors (e.g. programmed cell death 1/PD-1, PD1-Ligand/PDL-1) have been proved a breakthrough therapy for advanced HL [66, 73, 74].

The JAK/STAT pathway is activated in HL as a result of genomic amplification of JAK2 and/or inactivating mutations in an inhibitor of JAK activity, SOCS1 (75). A proof of the therapeutic potential of JAK inhibitors has been proved by a phase I study of the JAK inhibitor SB1518, a selective inhibitor of JAK2 and FLT3. In this study, 14 out of the 34 patients had cHL. Of these 14 patients, 6 patients had a steady disease with the treatment [31]. Have also supported the positive effects of SB1518, a novel macrocyclic pyrimidine-based JAK2 inhibitor for the treatment of HL. More specifically, SB1518 aims the JAK/STAT pathway by inhibiting tyrosine phosphorylation on JAK2 (Y221) and downstream STATs. Hence, SB1518 has probably an anti-proliferative effect on lymphoid cell lines, driven by mutant or wild type JAK-2 or FLT3. The latter results from cell cycle arrest and induction of apoptosis [31].

Furthermore, JAK inhibitors can lead on propitious immunomodulatory effects. Derenzini et al. [76] showed that AZD1480, JAK 1/2 inhibitor exhibited immunomodulatory effects at low concentrations by down regulating the expression of Th2 cytokines and chemokines (IL-13 and TRAC), as well as STAT3-mediated reduced expression of PD-L1 and PD-L2, which take part in immune escape mechanisms in HL. In addition. Demonstrated that JAK2 inhibition by the selective inhibitor, fedratinib decreased phosphorylation of JAK2, STAT1, STAT3, and STAT6 and reduced the expression of additional downstream targets, includ-

ing PD-L1. Interestingly, the phosphorylation of STAT 1, 3 and 6 was inhibited by chemical JAK2 blockade in a 9p24.1 copy number-dependent manner in cHL cell lines [20].

Recently presented a JAK 1 /2 inhibitor, ruxolitinib, which reduced the phosphorylation of STAT3 and STAT6, as well as the expression of c-Myc in the HL cell line HDLM-2. These results were amplified when ruxolitinib was combined with the Bcl-2/ Bcl-xL inhibitor, Navitoclax, or with anti-CD30 toxin conjugate, brentuximab vedotin (BV). The combination of ruxolitinib with Navitoclax or BV alone prolonged survival period but did not cure HDML-2 tumor-bearing mice. On the other, BV combined with ruxolitinib and/or with Navitoclax led to sustained complete remission in this model of HL. The studies above propose future use of the combination of BV with ruxolitinib in patients with HL [77].

The significance of STAT6 inhibition in HL therapy has been reported. The authors demonstrated a direct antiproliferative effect of histone deacetylase (HDAC) inhibitor vorinostat on HRS which was associated with cell cycle arrest and apoptosis and an immune mediated effect by altering cytokine and chemokines secretion in the microenvironment due to inhibition of STAT6 phosphorylation [78]. Furthermore demonstrated that the pan-deacetylase inhibitor panobinostat has potent antiproliferative activity against Hodgkin lymphoma-derived cell lines. At the molecular level, panobinostat activated the caspase pathway, inhibited STAT5 and STAT6 phosphorylation, and down-regulated hypoxia-inducible factor 1 α and its downstream targets, glucose transporter 1 (GLUT1) and vascular endothelial growth factor.

In respect to STAT6 signaling, have shown that specific blocking of the IL-4 and IL-13-mediated STAT-6 activation by an IL-4 binding fusion protein APG598 or an IL-4R antagonist APG201 (R121D/Y124D) make HL cells more prone to apoptotic effect by chemotherapeutic drugs such as Mitomycin C, 5-Fluoracil, Etoposide, Doxorubicin and Plaxicatel. This outcome is based on the inhibition of STAT-6 mediated elevation of expression of the anti-apoptotic Bcl-2 family protein Bcl-xL. Thus, the IL-4/ IL-13-STAT6-Bcl-xL pathway may be a crucial target for HL treatment [25]. In addition, Demonstrated that treatment of two IL-13-responsive HL-derived cell lines, with Soluble interleukin-13R α 2 decoy receptor, resulted in the inhibition of cell proliferation, and down-regulated the phosphorylation of STAT6 [24]. All the data investigating the correlation of STAT6 with cHL and its prognosis are summarized in **Table 1**.

Table1. Presentation of studies regarding STAT6 in Classical Hodgkin Lymphoma.

| Reference | STAT-6 related remarks | Association with prognosis |
|--|---|-------------------------------------|
| Skinnider <i>et al.</i> , 2002 | STAT-6 activation in cHL is mediated by IL-13 | Negative association with prognosis |
| Skinnider <i>et al.</i> , 2002, Hinz <i>et al.</i> , 2002 | STAT-6 is activated and expressed at high levels in HL | Negative association with prognosis |
| Kiss <i>et al.</i> , 2011 | STAT-6 plays an important role in the pathogenesis of EBV-positive cHL | Negative association with prognosis |
| Skinnider <i>et al.</i> , 2002 Natali <i>et al.</i> , 2013 Diaz <i>et al.</i> , 2011 | Survival and proliferation of HRS cells is dependent on STAT-6 and STAT-3 activation | Negative association with prognosis |
| Skinnider <i>et al.</i> , 2002 | Antibody-mediated neutralization of IL-13 reduced STAT-6 phosphorylation and decreased HL cell proliferation | Positive association with prognosis |
| Hart <i>et al.</i> , 20011 | JAK-STAT inhibitors seem to have a therapeutic potential | Positive association with prognosis |
| Hebenstreit <i>et al.</i> , 2016 | JAK2 selective inhibition decreases phosphorylation of STAT-6 and reduces the expression of downstream targets. | Positive association with prognosis |
| Buglio <i>et al.</i> , 2008 | Histone Deacetylase (HDAC) inhibitors decrease STAT-6 phosphorylation and promote cell cycle arrest and apoptosis | Positive association with prognosis |

6. Conclusion

In conclusion, although the complexity of interactions between HRS cells and their microenvironment and their functional role during malignant transformation is not completely understood, however, emerging data indicate that STAT6 is involved in cHL pathogenesis and growth, through interplay with cellular signal transduction pathways. Experimental results following disruption of microenvironmental interactions by STAT6 inhibition generate optimism for novel therapeutic strategies for HL, possibly including drugs that block specifically the STAT6 signaling pathway and particularly the STAT6 protein.

References

- Matsuki E, Younes A. Lymphomagenesis in Hodgkin lymphoma. *Semin Cancer Biol.* 2015;34:14-21.
- Zocchi MR, Catellani S, Canevali P, Tavella S, Garuti A, Villaggio B, et al. High ERp5/ADAM10 expression in lymph node microenvironment and impaired NKG2D ligands recognition in Hodgkin lymphomas. *Blood.* 2012;119(6):1479-89.
- Schmitz R, Stanelle J, Hansmann ML, Kuppers R. Pathogenesis of classical and lymphocyte-predominant Hodgkin lymphoma. *Annu Rev Pathol.* 2009;4:151-74.
- Kuppers R. The biology of Hodgkin's lymphoma. *Nat Rev Cancer.* 2009;9(1):15-27.
- Kuppers R, Engert A, Hansmann ML. Hodgkin lymphoma. *J Clin Invest.* 2012;122(10):3439-47.
- Liu Y, Sattarzadeh A, Diepstra A, Visser L, van den Berg A. The microenvironment in classical Hodgkin lymphoma: an actively shaped and essential tumor component. *Semin Cancer Biol.* 2014;24:15-22.
- Carbone A, Gloghini A, Caruso A, De Paoli P, Dolcetti R. The impact of EBV and HIV infection on the microenvironmental niche underlying Hodgkin lymphoma pathogenesis. *International journal of cancer.* 2016.
- Mathas S, Hartmann S, Kuppers R. Hodgkin lymphoma: Pathology and biology. *Semin Hematol.* 2016;53(3):139-47.
- Kilger E, Kieser A, Baumann M, Hammerschmidt W. Epstein-Barr virus-mediated B-cell proliferation is dependent upon latent membrane protein 1, which simulates an activated CD40 receptor. *Embo j.* 1998;17(6):1700-9.
- Kis LL, Gerasimcik N, Salamon D, Persson EK, Nagy N, Klein G, et al. STAT6 signaling pathway activated by the cytokines IL-4 and IL-13 induces expression of the Epstein-Barr virus-encoded protein LMP-1 in absence of EBNA-2: implications for the type II EBV latent gene expression in Hodgkin lymphoma. *Blood.* 2011;117(1):165-74.
- Shair KH, Bendt KM, Edwards RH, Bedford EC, Nielsen JN, Raab-Traub N. EBV latent membrane protein 1 activates Akt, NFkappaB, and Stat3 in B cell lymphomas. *PLoS Pathog.* 2007;3(11):e166.
- Warnke RA WL, Chan JK et al. . Classic Hodgkin's disease. In Atlas of tumor Pathology. Series III, fascicle Tumors of the lymph nodes and spleen. Atlas of tumor pathology 1995:277-304.
- Skinnider BF, Kapp U, Mak TW. Interleukin 13: a growth factor in hodgkin lymphoma. *Int Arch Allergy Immunol.* 2001;126(4):267-76.
- Skinnider BF, Mak TW. The role of cytokines in classical Hodgkin lymphoma. *Blood.* 2002;99(12):4283-97.
- Wolf J, Kapp U, Bohlen H, Kornacker M, Schoch C, Stahl B, et al. Peripheral blood mononuclear cells of a patient with advanced Hodgkin's lymphoma give rise to permanently growing Hodgkin-Reed Sternberg cells. *Blood.* 1996;87(8):3418-28.
- Klein S, Jucker M, Diehl V, Tesch H. Production of multiple cytokines by Hodgkin's disease derived cell lines. *Hematol Oncol.* 1992;10(6):319-29.
- Dukers DE, Jaspars LH, Vos W, Oudejans JJ, Hayes D, Cillessen S, et al. Quantitative immunohistochemical analysis of cytokine profiles in Epstein-Barr virus-positive and -negative cases of Hodgkin's disease. *J Pathol.* 2000;190(2):143-9.
- Warnke RA WL CJ. Tumors of the lymph nodes and spleen. Classic Hodgkin's disease. 1995.
- Godava M, Vrtel R, Vodicka R. STAT6 - polymorphisms, haplotypes and epistasis in relation to atopy and asthma. *Biomed Pap Med Fac Univ*

- Palacky Olomouc Czech Repub. 2013;157(2):172-80.
20. Hebenstreit D, Wirnsberger G, Horejs-Hoeck J, Duschl A. Signaling mechanisms, interaction partners, and target genes of STAT6. *Cytokine Growth Factor Rev.* 2006;17(3):173-88.
 21. Skinnider BF, Elia AJ, Gascoyne RD, Patterson B, Trumper L, Kapp U, et al. Signal transducer and activator of transcription 6 is frequently activated in Hodgkin and Reed-Sternberg cells of Hodgkin lymphoma. *Blood.* 2002;99(2):618-26.
 22. Hao Y, Chapuy B, Monti S, Sun HH, Rodig SJ, Shipp MA. Selective JAK2 inhibition specifically decreases Hodgkin lymphoma and mediastinal large B-cell lymphoma growth in vitro and in vivo. *Clin Cancer Res.* 2014;20(10):2674-83.
 23. Raia V, Schilling M, Bohm M, Hahn B, Kowarsch A, Raue A, et al. Dynamic mathematical modeling of IL13-induced signaling in Hodgkin and primary mediastinal B-cell lymphoma allows prediction of therapeutic targets. *Cancer Res.* 2011;71(3):693-704.
 24. Trieu Y, Wen XY, Skinnider BF, Bray MR, Li Z, Claudio JO, et al. Soluble interleukin-13Ralpha2 decoy receptor inhibits Hodgkin's lymphoma growth in vitro and in vivo. *Cancer Res.* 2004;64(9):3271-5.
 25. Natoli A, Lupertz R, Merz C, Muller WW, Kohler R, Krammer PH, et al. Targeting the IL-4/IL-13 signaling pathway sensitizes Hodgkin lymphoma cells to chemotherapeutic drugs. *Int J Cancer.* 2013;133(8):1945-54.
 26. Batlevi CL, Younes A. Novel therapy for Hodgkin lymphoma. *Hematology Am Soc Hematol Educ Program.* 2013;2013:394-9.
 27. Lemoine M, Derenzini E, Buglio D, Medeiros LJ, Davis RE, Zhang J, et al. The pan-deacetylase inhibitor panobinostat induces cell death and synergizes with everolimus in Hodgkin lymphoma cell lines. *Blood.* 2012;119(17):4017-25.
 28. Blum KA, Johnson JL, Niedzwiecki D, Canellos GP, Cheson BD, Bartlett NL. Single agent bortezomib in the treatment of relapsed and refractory Hodgkin lymphoma: cancer and leukemia Group B protocol 50206. *Leuk Lymphoma.* 2007;48(7):1313-9.
 29. Younes A, Pro B, Fayad L. Experience with bortezomib for the treatment of patients with relapsed classical Hodgkin lymphoma. *Blood.* 2006;107(4):1731-2.
 30. Fanale M, Fayad L, Pro B, Samaniego F, Liboon MJ, Nunez C, et al. Phase I study of bortezomib plus ICE (BICE) for the treatment of relapsed/refractory Hodgkin lymphoma. *Br J Haematol.* 2011;154(2):284-6.
 31. Hart S, Goh KC, Novotny-Diermayr V, Hu CY, Hentze H, Tan YC, et al. SB1518, a novel macrocyclic pyrimidine-based JAK2 inhibitor for the treatment of myeloid and lymphoid malignancies. *Leukemia.* 2011;25(11):1751-9.
 32. Kaplan MH, Schindler U, Smiley ST, Grusby MJ. Stat6 is required for mediating responses to IL-4 and for development of Th2 cells. *Immunity.* 1996;4(3):313-9.
 33. Shimoda K, van Deursen J, Sangster MY, Sarawar SR, Carson RT, Tripp RA, et al. Lack of IL-4-induced Th2 response and IgE class switching in mice with disrupted Stat6 gene. *Nature.* 1996;380(6575):630-3.
 34. Takeda K, Tanaka T, Shi W, Matsumoto M, Minami M, Kawashima S, et al. Essential role of Stat6 in IL-4 signalling. *Nature.* 1996;380(6575):627-30.
 35. Wurster AL, Tanaka T, Grusby MJ. The biology of Stat4 and Stat6. *Oncogene.* 2000;19(21):2577-84.
 36. Goenka S, Kaplan MH. Transcriptional regulation by STAT6. *Immunol Res.* 2011;50(1):87-96.
 37. Walford HH, Doherty TA. STAT6 and lung inflammation. *Jakstat.* 2013;2(4):e25301.
 38. Hershey GK. IL-13 receptors and signaling pathways: an evolving web. *J Allergy Clin Immunol.* 2003;111(4):677-90; quiz 91.
 39. Zimmermann N, Hershey GK, Foster PS, Rothenberg ME. Chemokines in asthma: cooperative interaction between chemokines and IL-13. *J Allergy Clin Immunol.* 2003;111(2):227-42; quiz 43.
 40. Ni Z, Lou W, Lee SO, Dhir R, DeMiguel F, Grandis JR, et al. Selective activation of members of the signal transducers and activators of transcription family in prostate carcinoma. *J Urol.* 2002;167(4):1859-62.
 41. Zhang MS, Zhou YF, Zhang WJ, Zhang XL, Pan Q, Ji XM, et al. Apoptosis induced by short hairpin RNA-mediated STAT6 gene silencing in human colon cancer cells. *Chin Med J (Engl).* 2006;119(10):801-8.
 42. Li BH, Yang XZ, Li PD, Yuan Q, Liu XH, Yuan J, et al. IL-4/Stat6 activities correlate with apoptosis and metastasis in colon cancer cells. *Biochem Biophys Res Commun.* 2008;369(2):554-60.
 43. Merk BC, Owens JL, Lopes MB, Silva CM, Hussaini IM. STAT6 expression in glioblastoma promotes invasive growth. *BMC Cancer.* 2011;11:184.
 44. Benekli M, Baer MR, Baumann H, Wetzler M. Signal transducer and activator of transcription proteins in leukemias. *Blood.* 2003;101(8):2940-54.
 45. Guiter C, Dusanter-Fourt I, Copie-Bergman C, Boulland ML, Le Gouvello S, Gaulard P, et al. Constitutive STAT6 activation in primary mediastinal large B-cell lymphoma. *Blood.* 2004;104(2):543-9.
 46. Melzner I, Bucur AJ, Bruderlein S, Dorsch K, Hasel C, Barth TF, et al. Biallelic mutation of SOCS-1 impairs JAK2 degradation and sustains phospho-JAK2 action in the MedB-1 mediastinal lymphoma line. *Blood.* 2005;105(6):2535-42.
 47. Bruns HA, Kaplan MH. The role of constitutively active Stat6 in leukemia and lymphoma. *Crit Rev Oncol Hematol.* 2006;57(3):245-53.

48. Robinson DR, Wu YM, Kalyana-Sundaram S, Cao X, Lonigro RJ, Sung YS, et al. Identification of recurrent NAB2-STAT6 gene fusions in solitary fibrous tumor by integrative sequencing. *Nat Genet.* 2013;45(2):180-5.
49. Chmielecki J, Crago AM, Rosenberg M, O'Connor R, Walker SR, Ambrogio L, et al. Whole-exome sequencing identifies a recurrent NAB2-STAT6 fusion in solitary fibrous tumors. *Nat Genet.* 2013;45(2):131-2.
50. Doyle LA, Vivero M, Fletcher CD, Mertens F, Hornick JL. Nuclear expression of STAT6 distinguishes solitary fibrous tumor from histologic mimics. *Mod Pathol.* 2014;27(3):390-5.
51. Vogels RJ, Vlenterie M, Versleijen-Jonkers YM, Ruijter E, Bekers EM, Verdijk MA, et al. Solitary fibrous tumor - clinicopathologic, immunohistochemical and molecular analysis of 28 cases. *Diagn Pathol.* 2014;9:224.
52. Doyle LA, Tao D, Marino-Enriquez A. STAT6 is amplified in a subset of dedifferentiated liposarcoma. *Mod Pathol.* 2014;27(9):1231-7.
53. Feys T, Poppe B, De Preter K, Van Roy N, Verhasselt B, De Paepe P, et al. A detailed inventory of DNA copy number alterations in four commonly used Hodgkin's lymphoma cell lines. *Haematologica.* 2007;92(7):913-20.
54. Furqan M, Akinleye A, Mukhi N, Mittal V, Chen Y, Liu D. STAT inhibitors for cancer therapy. *J Hematol Oncol.* 2013;6:90.
55. Oh CK, Geba GP, Molfino N. Investigational therapeutics targeting the IL-4/IL-13/STAT-6 pathway for the treatment of asthma. *Eur Respir Rev.* 2010;19(115):46-54.
56. Tiacchi E, Doring C, Brune V, van Noesel CJ, Klapper W, Mechttersheimer G, et al. Analyzing primary Hodgkin and Reed-Sternberg cells to capture the molecular and cellular pathogenesis of classical Hodgkin lymphoma. *Blood.* 2012;120(23):4609-20.
57. Kreher S, Bouhlel MA, Cauchy P, Lamprecht B, Li S, Grau M, et al. Mapping of transcription factor motifs in active chromatin identifies IRF5 as key regulator in classical Hodgkin lymphoma. *Proc Natl Acad Sci U S A.* 2014;111(42):E4513-22.
58. Hinz M, Lemke P, Anagnostopoulos I, Hacker C, Krappmann D, Mathas S, et al. Nuclear factor kappaB-dependent gene expression profiling of Hodgkin's disease tumor cells, pathogenetic significance, and link to constitutive signal transducer and activator of transcription 5a activity. *J Exp Med.* 2002;196(5):605-17.
59. Johnson PC, McAulay KA, Montgomery D, Lake A, Shield L, Gallagher A, et al. Modeling HLA associations with EBV-positive and -negative Hodgkin lymphoma suggests distinct mechanisms in disease pathogenesis. *Int J Cancer.* 2015;137(5):1066-75.
60. Pantanowitz L, Carbone A, Dolcetti R. Microenvironment and HIV-related lymphomagenesis. *Semin Cancer Biol.* 2015; 34: 52-7.
61. Kapatai G, Murray P. Contribution of the Epstein Barr virus to the molecular pathogenesis of Hodgkin lymphoma. *J Clin Pathol.* 2007; 60(12): 1342-9.
62. Middeldorp JM, Pegtel DM. Multiple roles of LMP1 in Epstein-Barr virus induced immune escape. *Semin Cancer Biol.* 2008; 18(6): 388-96.
63. Hannigan A, Wilson JB. Evaluation of LMP1 of Epstein-Barr virus as a therapeutic target by its inhibition. *Mol Cancer.* 2010; 9: 184.
64. Diaz T, Navarro A, Ferrer G, Gel B, Gaya A, Artells R. Lestaurtinib inhibition of the Jak/STAT signaling pathway in Hodgkin lymphoma inhibits proliferation and induces apoptosis. *PLoS One.* 2011; 6(4): e18856.
65. Green MR, Monti S, Rodig SJ, Juszczynski P, Currie T, O'Donnell E, et al. Integrative analysis reveals selective 9p24.1 amplification, increased PD-1 ligand expression, and further induction via JAK2 in nodular sclerosing Hodgkin lymphoma and primary mediastinal large B-cell lymphoma. *Blood.* 2010; 116(17): 3268-77.
66. Ansell SM, Lesokhin AM, Borrello I, Halwani A, Scott EC, Gutierrez M, et al. PD-1 blockade with nivolumab in relapsed or refractory Hodgkin's lymphoma. *N Engl J Med.* 2015; 372(4): 311-9.
67. Juszczynski P, Ouyang J, Monti S, Rodig SJ, Takeyama K, Abramson J, et al. The API-1-dependent secretion of galectin-1 by Reed Sternberg cells fosters immune privilege in classical Hodgkin lymphoma. *Proc Natl Acad Sci U S A.* 2007; 104(32): 13134-9.
68. Skinnider BF, Elia AJ, Gascoyne RD, Trumper LH, von Bonin F, Kapp U, et al. Interleukin 13 and interleukin 13 receptor are frequently expressed by Hodgkin and Reed-Sternberg cells of Hodgkin lymphoma. *Blood.* 2001; 97(1): 250-5.
69. Skinnider BF, Kapp U, Mak TW. The role of interleukin 13 in classical Hodgkin lymphoma. *Leuk Lymphoma.* 2002; 43(6): 1203-10.
70. Aldinucci D, Celegato M, Casagrande N. Microenvironmental interactions in classical Hodgkin lymphoma and their role in promoting tumor growth, immune escape and drug resistance. *Cancer Lett.* 2016; 380(1): 243-52.
71. Brice P. Managing relapsed and refractory Hodgkin lymphoma. *Br J Haematol.* 2008; 141(1): 3-13.
72. Younes A. Novel treatment strategies for patients with relapsed classical Hodgkin lymphoma. *Blood Rev.* 2010; 24(6): 233-8.
73. Villasboas JC, Ansell S. Checkpoint Inhibition: Programmed Cell Death 1 and Programmed Cell Death 1 Ligand Inhibitors in Hodgkin Lymphoma. *Cancer J.* 2016; 22(1): 17-22.
74. Carlo-Stella C, Santoro A. Microenvironment-related biomarkers and novel targets in classical Hodgkin's lymphoma. *Biomark Med.* 2015; 9(8): 807-17.
75. Kuppers R. New insights in the biology of Hodgkin lymphoma. *He-*

matology Am Soc Hematol Educ Program. 2012; 2012: 328-34.

76. Derenzini E, Lemoine M, Buglio D, Katayama H, Ji Y, Davis RE, et al. The JAK inhibitor AZD1480 regulates proliferation and immunity in Hodgkin lymphoma. *Blood Cancer J*. 2011; 1(12):e46.

77. Ju W, Zhang M, Wilson KM, Petrus MN, Bamford RN, Zhang X, et al. Augmented efficacy of brentuximab vedotin combined with ruxolitinib and/or Navitoclax in a murine model of human Hodgkin's lymphoma. *Proc Natl Acad Sci U S A*. 2016; 113(6): 1624-9.

78. Buglio D, Georgakis GV, Hanabuchi S, Arima K, Khaskhely NM, Liu YJ, et al. Vorinostat inhibits STAT6-mediated TH2 cytokine and TARC production and induces cell death in Hodgkin lymphoma cell lines. *Blood*. 2008;112(4): 1424-33.